

CHRIST COLLEGE (AUTONOMOUS), IRINJALAKUDA



DEGREE OF M. Sc. Botany

MASTER OF SCIENCE IN BOTANY

**(CHOICE BASED CREDIT AND SEMESTER SYSTEM FOR
POSTGRADUATE CURRICULUM)**

UNDER THE FACULTY OF SCIENCE

SYLLABUS

(FOR THE STUDENTS ADMITTED FROM THE ACADEMIC YEAR 2019 – '20 ONWARDS)

BOARD OF STUDIES IN BOTANY (PG)

CHRIST COLLEGE (AUTONOMOUS), IRINJALAKUDA - 680125, KERALA, INDIA

JUNE, 2019

CHRIST COLLEGE (AUTONOMOUS), IRINJALAKUDA

M. Sc. Programme in Botany (CBCSS) (from 2019 admissions onwards)

Programme structure of courses and distribution of credits

Course	Title	Credits		
		Internal	External	Total credits
Semester I				
BOT1C01	Phycology, Bryology, Pteridology and Gymnosperms	20%	80%	5
BOT1C02	Mycology and Lichenology, Microbiology and Plant Pathology	20%	80%	5
BOT1C03	Angiosperm Anatomy, Angiosperm Embryology, Palynology and Lab Techniques	20%	80%	5
BOT1L01	Practicals of Phycology, Bryology, Pteridology, Gymnosperms, Mycology and Lichenology	20%	80%	2.5
BOT1L02	Practicals of Microbiology, Plant Pathology, Angiosperm Anatomy, Angiosperm Embryology, Palynology and Lab Techniques.	20%	80%	2.5
Semester II				
BOT2C04	Cell Biology, Molecular Biology and Biophysics	20%	80%	5
BOT2C05	Cytogenetics, Genetics, Biostatistics, Plant Breeding and Evolution	20%	80%	5
BOT2C06	Plant Ecology, Conservation Biology, Phytogeography and Forest Botany	20%	80%	5
BOT2L03	Practicals of Cell Biology, Molecular Biology, Biophysics and Cytogenetics	20%	80%	2.5
BOT2L04	Practicals of Genetics, Biostatistics, Plant Breeding, Plant Ecology, Conservation Biology, Phytogeography and Forest Botany	20%	80%	2.5
Semester III				
BOT3C07	Plant Physiology, Metabolism and Biochemistry	20%	80%	5
BOT3C08	Angiosperm Morphology, Angiosperm Taxonomy and Plant Resources	20%	80%	5
BOT3C09	Biotechnology and Bioinformatics	20%	80%	5
BOT3L05	Practicals of Plant Physiology, Metabolism, Biochemistry, Angiosperm Morphology and Angiosperm Taxonomy	20%	80%	2.5
BOT3L06	Practicals of Plant Resources, Biotechnology and Bioinformatics	20%	80%	2.5
Semester IV				
BOT4E01	Elective I	20%	80%	5
BOT4E02	Elective II	20%	80%	5
BOT4L07	Practicals of Electives	20%	80%	2
BOT4D01	Dissertation	20%	80%	5
BOT4V01	Viva voce	0%	100%	3
Total				80 credits
Audit Courses (To be completed within the first three semesters by the students)				
ACIAEC	Ability Enhancement Course	100%	0%	4
AC2PCC	Professional Competency Course	100%	0%	4
(The credits earned through the audit courses will not be added for SGPA/CGPA)				
Duration of Theory Examinations (External) as well as Practical Examinations (External) will be 3 hours				
1 credit = 1.25 hours of teaching; There will be no regular classes/workload for audit courses.				
1 theory/dissertation hour= 1 hours of workload; 1 practical hour= 1 hour of workload				

ADMISSION:

Admission for the Programme shall be as per the CBCSS PG Regulations in force.

ATTENDANCE:

The requirement of attendance shall be as per the CBCSS PG Regulation in force.

EVALUATION AND GRADING

EVALUATION: The evaluation scheme for each course shall contain two parts; (a) Internal / Continuous Assessment (CA) and (b) External / End Semester Evaluation (ESE). Of the total, 20% weightage shall be given to Internal evaluation / Continuous assessment and the remaining 80% to External/ESE and the ratio and weightage between Internal and External is 1:4. Primary evaluation for Internal and External shall be based on 6 letter grades (A+, A, B, C, D and E) with numerical values (Grade Points) of 5, 4, 3, 2, 1 & 0 respectively.

Grade Point Average: Internal and External components are separately graded and the combined grade point with weightage 1 for Internal and 4 for external shall be applied to calculate the Grade Point Average (GPA) of each course. Letter grade shall be assigned to each course based on the categorization based on Ten-point scale.

Evaluation of Audit Courses: The examination and evaluation shall be conducted by the college in a common pattern for all the PG programs. The question paper shall be for minimum 20 weightage and a minimum of 2-hour duration for the examination. The result has to be intimated/ uploaded to the University during the Third Semester as per the notification of the University.

INTERNAL EVALUATION / CONTINUOUS ASSESSMENT (CA)

This assessment shall be based on a predetermined transparent system involving periodic written tests, assignments, seminars and viva-voce in respect of theory courses and based on tests, lab skill and records/viva in respect of practical courses.

The criteria and percentage of weightage assigned to various components for internal evaluation are as follows:

(a) Theory:		Component	Percentage Weightage
SI. No.			
1		Examination /Test	40%
2		Seminars / Presentation	20%
	3	Assignment	20%
	4	Attendance	20%
(b) Practical:			
1		Lab Skill	40%
2		Records	30%
3		Practical Test	30%

(Grades shall be given for the internal evaluation based on the grades A+, A, B, C, D & E with grade points 5,4,3,2, 1 & 0 respectively. The overall grades shall be as per the Ten-point scale. There shall be no separate minimum Grade Point for internal evaluation. To ensure transparency of the evaluation process, the internal assessment marks awarded to the students in each course in a semester shall be published on the notice board before 5 days of commencement of external examination. There shall not be any chance for improvement of internal marks. The course teacher shall maintain the academic record of each student registered for the course. Class tests for internal evaluation should be spread during the semester and the grades displayed on the notice board. Valued answer scripts shall be made available to the students for perusal. Each student shall be required to do at least one assignment for each course. Assignments after valuation must be returned to the students. The teacher shall define the expected quality of the above in terms of structure, content, presentation etc. and inform the same to the students. Punctuality in submission is to be considered. Every student shall deliver one seminar / presentation as an internal component for every course and must be evaluated by the respective course teacher in terms of structure, content, presentation and interaction. The soft and hard copies of the seminar report are to be submitted to the course teacher. All the records of Continuous Assessment (CA) must be kept in the college and must be made available for verification, if asked for.)

EXTERNAL / END SEMESTER EVALUATION (ESE)

The semester-end examinations in theory courses shall be conducted by the College with question papers set by external experts. The evaluation of the answer scripts shall be done by examiners based on a well-defined scheme of valuation. After the external evaluation, only Grades are to be entered in the space provided in the answer script for individual questions and calculations need to be done only up to the Cumulative Grade Point (CGP). Students shall have the right to apply for revaluation or scrutiny as per rules within the time permitted for it. Photocopies of the answer scripts of the external examination shall be made available to the students for scrutiny on request by them as per rules. At the end of each semester, there will be external evaluation for each course. The theory examination will be of 3-hour duration and will have a total of 30 weightage. There will be two evaluations for the theory papers - one by an internal examiner and the other by an external examiner.

PATTERN OF QUESTIONS FOR EXTERNAL ESE

Questions shall be set to assess the knowledge acquired, standard, and application of knowledge, application of knowledge in new situations, critical evaluation of knowledge and the ability to synthesize knowledge. Due weightage shall be given to each module based on content/teaching hours allotted to each module. It has to be ensured that questions covering all skills are set. The setter shall also submit a detailed scheme of evaluation along with the question paper. A question paper shall be a judicious mix of short answer type, short essay type /problem solving type and long essay type questions. The question shall be prepared in such a way that the answers can be awarded A+, A, B, C, D & E Grades. End Semester Evaluation in Practical Courses shall be conducted and evaluated by two examiners of which one should be an External Examiner and the other examiner should be the teacher who offers the course/ the senior most teacher who offers the course. Different types of questions shall be given different weightages to quantify their range given in the following model:

Number of questions to be answered:**1. Theory**

Sl. No.	Type of Individual Questions	Total No. of Questions	Weightage
1.	Short answer	4 out of 7	2x4=8
2.	Short essay/problem solving	4 out of 7	3x4=12
3	Long Essay type	2 out of 4	5x2=10
	Total	10 out of 18	30

(All questions should be in such a way that 6 grades could be awarded. Short answer questions should have a minimum of 4 value points, short essays a minimum of 6 value points and long essays a minimum of 10 value points)

2. Practicals

Sl. No.	Type of Individual Questions	Total No. of Questions	Weightage
1.	Major Experiments/ Problems	3	3x5=15
2.	Minor Experiments/ Problems	3	3x2=6
3	Spotters/ Identifications	5	5x1=5
4	Lab Records	1	2
5	Submissions/Tour Reports	1	2
	Total	13	30

EVALUATION OF PROJECT WORK | DISSERTATION

There shall be External and Internal evaluation with the same criteria for Project Work done and the grading system shall be followed as per the specific guidelines. For a pass in Project Work, a student has to secure a minimum of P Grade in External and Internal examination combined. If the students could not secure minimum P Grade in the Project work, they will be treated as failed in that attempt and the students may be allowed to rework and resubmit the same in accordance with the Institution exam stipulations. There shall be no improvement chance for Project Work.

The External and Internal evaluation of the Project Work shall be done based on the following criteria and weightages as detailed below:

Sl. No.	Criteria	% of Weightage	Weightage
1	Relevance of the topic	10	3
2	Methodology & Analysis	40	12
3	Discussion	10	3
4	Viva Voce (on the project)	40	12
	Total	100	30

COMPREHENSIVE VIVA-VOCE

There shall be an External Comprehensive Viva-Voce at the end of the IVth Semester. The External Viva Voce shall be conducted by one External Examiner from an outside institution and the Head of the Department as the Internal Examiner. For a pass in comprehensive viva-voce, a student has to secure a minimum of P Grade or a pass. Failed candidates can reappear for the same next time in accordance with the Institution exam stipulations. There shall be no improvement chance for comprehensive viva-voce.

DIRECT GRADING SYSTEM

Direct Grading System based on a 10 Point scale is used to evaluate the performance (External and Internal Examinations of students) for all courses (Theory & Practical)/Semester/Overall Programme, Letter grades and GPA/SGPA/CGPA are given on the following way:

a) First Stage Evaluation for both Internal and External will be done by the teachers concerned in the following

scale:	A+	5
	A	4
	B	3
	C	2
	D	1
	E	0

b) The Grade Range for both Internal & External shall be:

Letter Grade	Grade Range	Range of %	Merit indicator
O	4.25 - 5.00	85-100	Outstanding
A+	3.75 - 4.24	75-84.99	Excellent
A	3.25 - 3.74	65-74.99	Very Good
B+	2.75 - 3.24	55-64.99	Good
B	2.50 - 2.74	50-54.99	Above Average
C	2.25 - 2.49	45-49.99	Average
P	2.00 - 2.24	40-44.99	Pass
F	< 2.00	Below 40	Fail
I	0		Incomplete
Ab	0		Absent

No separate minimum is required for Internal Evaluation for a pass, but a minimum P Grade is required for a pass in the external evaluation. However, a minimum P grade is required for pass in a course. A student who fails to secure a minimum grade for a pass in a course will be permitted to write the examination along with the next batch.

IMPROVEMENT OF COURSE

The candidates who wish to improve the grade / grade point of the external examination of a course they have passed already can do the same by appearing in the external examination of the concerned semester along with the immediate junior batch. A candidate will be permitted to improve the CGPA of the Programme within a continuous period of four semesters immediately following the completion of the programme allowing only once for a particular semester. The CGPA for the betterment appearance will be computed based on the SGPA secured in the original or betterment appearance of each semester whichever is higher.

SGPA CALCULATION

SGPA is the ratio of sum of the product of the number of credits with the grade points scored by a student in all the courses taken by a student and the sum of the number of credits of all the courses taken by a student. After the successful completion of a semester, Semester Grade Point Average (SGPA) of a student in that semester is calculated using the formula given below:

$$\text{SGPA (S}_j\text{)} = \Sigma(\text{C}_i \times \text{G}_i) / \text{Cr}$$

Where 'S_j' is the j semester, 'G_i' is the grade point scored by the student in the i course 'c_i' is the credit of the i course, 'Cr' is the total credits of the semester.

CGPA CALCULATION

$$\text{CGPA} = \Sigma(\text{C}_i \times \text{S}_i) / \text{Cr}$$

Where C_i is the credit of the ith semester, S_i is the SGPA of the ith semester and Cr is the total number of credits in the programme. The CGPA is also calculated in the same manner taking into account all the courses undergone by a student over all the semesters of a programme. The SGPA and CGPA shall be rounded off to 2 decimal points. For the successful completion of a semester, a student should pass all courses and score a minimum SGPA of 2.0. However, the students are permitted to move to the next semester irrespective of their SGPA.

SEMESTER I

BOT1C01 – PHYCOLOGY, BRYOLOGY, PTERIDOLOGY GYMNOSPERMS

Contact Hours per Week: 6

Course Outline

Phycology

1. Classification of Algae-comparative Survey of important systems - Fritsch-Smith-Round. Criteria for algal classification-Phylogenetic considerations.
2. Biological importance of Planktons.
3. Algal cytology-Basic ideas of cell features-Electron microscopic studies of algal cell, cell wall, flagella, chloroplast, pyrenoid, eyespot- their importance in classification.
4. Reproduction-Different types of life cycles in algae.
5. General account of energy sources and pigments in algae.
6. Economic importance of algae-Roll of algae in soil fertility, algae in industry-Biological importance of phytoplanktons and water blooms.
7. General account of thallus structure, cell ultra-structure, reproduction, relationships and evolutionary trends in the following' groups: Chlorophyta, Xanthophyta, Bacillariophyta, Phaeophyta, Rhodophyta.

References

1. Fritsch, F.E. The structure and Reproduction of Algae.
2. Smith, G.M. Manual of Phycology
3. Round, F.E, The Biology of Algae.
4. Pold and Wyane. Introduction of Algae.

Bryology

1. General characters and systems of classifications of Bryophytes
2. General account of the anatomy, reproduction, life history and phylogeny of Sphaerocarpaceales, Marchantiales, Jungermanniales, Calobryales, Anthocerotales, Sphagnales, Andreales, Funariales and Polytrichales
3. Origin and evolution of Bryophytes- gametophytic and sporophytic.
4. A general account of fossil Bryophytes and their affinities.
5. Economic importance of Bryophytes.

References

1. Watson E.V. The structure and life of Bryophytes. Hutchinson Univ. Press, London.
2. Cavers F. The interrelationship of Bryophytes. New Phytologist.
3. Kashyap S. R., The Liverworts of Western Himalaya and the Punjab Plains, Vol. I & II. Chronica Botanica
4. Smith G. M. Cryptogamic Botany. McGraw Hill Book Co., N.Y.
5. Parihar N. S. An introduction of Embryophyta: Bryophyta. General Book House, Allahabad.
6. Verdoon, F. M. Manual of Bryology. Ashor & Co., Amsterdam.
7. Shaw, J. and Goffinet, B. Bryophyte Biology. Cambridge University Press.
8. Manju C. Nair, K.P. Rajesh and Madhusoodanan P.V. Bryophytes of Wayanad in Western Ghats. Malabar Natural History Society, Kozhikode.

Pteridology

1. General characters and life history of Pteridophytes.
2. Cytology of Pteridophytes- Chromosome number and polyploidy.
3. Structure and evolution of stele in Pteridophytes.
4. Origin and evolution of Sporangium.
5. Heterospory and seed habit.
6. Development and evolutionary trends in the Gametophytes of Pteridophytes.
7. Apogamy, Apospory and Parthenogenesis.
8. Classification of Pteridophytes: Holttum, Pichi-Sermolli.
9. Comparative morphology, ecology and phylogeny of the following:
 - a) Psilopsida : Rhyniales, Psilophytales and Psilotales
 - b) Lycopsidea: Lycopodiales and Isoetales
 - c) Sphenopsida: Hyeniales, Pseudobomiales, Sphenophyllales, Calamitales and Equisetales.
 - d) Filicopsida: General account: Primofilicales, Ophioglossales, Marattiales, Osmundales, Schizaeales, Cyatheales, Gleicheniales, Marsileales and Salviniiales.
10. Economic importance of Pteridophytes-Medicinal, Horticulture, Biofertilizer, weeds.
11. General account of the contribution of Indian pteridologists.

References

1. Bierhost, D.W. Morphology of Vascular Plants. Mac Miilan Co., New York.
2. Dyer, A.C. The Experimental Biology of Ferns. Academic Press, London.
3. Jermy, A.C. (Ed.): The phylogeny and Classification of Ferns.
4. Kramer, K.U. and Green, P.S. The Families and Genera of Vascular Plants. Narosa, New Delhi.
5. Nampy, S. and Madhusoodanan, P.V. Fern Flora of South India-Taxonomic Revision of Polypodioid Ferns. Dava Publishing House, New Delhi.

6. Abdul Hameed C., Rajesh K.P. and Madhusoodanan P.V. *Filmy Ferns of South India*. Penta Book Publishers & Distributors, Calicut.
7. Azeez K., Venugopalakrishna Kurup V. and P.V. Madhusoodanan. *Spleenworts (Aspleniaceae) of South India*. Malabar Natural History Society, Calicut.
8. Venugopalakrishna Kurup V., Azeez K. and P.V. Madhusoodanan. *Primitive Ferns of South India*. 'V'Publishers, Kottayam.

Gymnosperms

1. Geological time scale and correlated predominant Gymnosperm flora. Classification of Gymnosperms- Chamberlain's system.
2. Geological horizons. Distribution, morphology, anatomy, reproduction and interrelationship of the following orders (Study of families and types not required)
 - a. Pteridospermales; b. Glossopteridales; c. Caytoniales; d. Cycadaeoidales; e. Pentoxylales; f. Cycadales, g. Ginkgoales; h. Cordaitales; i. Coniferales; j. Taxales; k. Ephedrales; l. Welwitschiales; m. Gnetales
3. Phylogenetic relationship of Gymnosperms.
4. Economic importance of Gymnosperms

References

1. Andrews, H.N. *Studies in Paleobotany*, Wiley, N.Y.
2. Banks, H.P. *Evolution and plants of the past*. Wadsworth.
3. Bierhost, D.W. *Morphology of Vascular Plants*. Macmillan.
4. Bower, F.O. *Primitive Plants*. Macmillan.
5. Chamberlain, C.J. *Gymnosperms- Structure and Evolution*. Univ. of Chicago Press.
6. Foster, A.S. & E.M. Gifford. *Comparative morphology of vascular plants*. Freeman.
7. Maheshwari, P & V. Vasil. *Gnetum*. CSIR, New Delhi.
8. Ramanujam, C.G.K. *Indian Gymnosperms in time and space*. Today & Tomorrow, Dehra Dun.
9. Sewart, W.N. *Paleobotany and the Evolution of Plants*. Cambridge Univ. Press.
10. Stockey, R.S. Some comments on the origin and evolution of conifers. *Canadian J. Bot.* 59: 75-82.
11. Taylor, T.N. Reproductive biology in early seed plants. *Bioscience* 32:23-28.
12. Walton. *An Introduction to the Study of Fossil plants*.

BOT1C02 – MYCOLOGY, LICHENOLOGY, MICROBIOLOGY AND PLANT PATHOLOGY

Contact Hours per Week: 6

Course Outline

Mycology

1. General characters of Fungi: cell-ultra structure, unicellular and multicellular organization, hyphal growth, cell wall composition, nutrition (saprobic, biotrophic, symbiotic, predacious) reproduction (vegetative, asexual, sexual), heterothallism, parasexuality.
2. Classification of fungi by Ainsworth & Bisby (1983), Alexopoulos et al. (1996)- Phylogeny of fungi- Characters used in classification.
3. General account of Myxomycota, Mastigomycota, Zygomycota, Ascomycota, Basidiomycota and mitosporic fungi. Different kinds of spores and their dispersal.
4. Fungi as saprophytes: details of the fungal decomposition of organic matter, coprophilous fungi, lignin degrading fungi, role of fungi in degradation of pesticides.
5. Fungi as symbionts: Mycorrhiza – ectotrophic, orchidaceous and Ericoid mycorrhiza, Vesicular Arbuscular Mycorrhiza - their distribution and significance. Endophytes.
6. Lichenology: General account and systematics of lichens, thallus structure, reproductive bodies, ecological significance and economic importance of lichens.

References

1. Alexopoulos C.J., Mims, C.W. & Blackwell, M. Introductory Mycology. 4th edition. John Wiley & Sons Inc.
2. Ainsworth, G.C., Sparrow, K.F.& Susmann, A.S.(Eds.). The Fungi - An Advanced Treatise. Vol 1-4. Academic Press.
3. Burnett, J.H. Fundamentals of Mycology. Edward Arnolds.
4. Cariile, M. J. & Watkinson S.C. The Fungi. Academic Press.
5. Deacon, J.W. Introduction to Modern Mycology. Blackwell.
6. Dubey, H.C. An Introduction to Fungi. Vikas Publishers, New Delhi.
7. Hale Mason, E. The Biology of Lichens. 3rd Ed. Edward Arnold, London.
8. Jennigs, D.H. & Lysek, G. Fungal Biology. Bios Scientific Publishers.
9. Mehrotra, R.S. & Aneja, K.R. An Introduction to Mycology. New Age International Publishers.
10. Landecker, Elizabeth Moore. Fundamentals of Fungi. 4th Ed. Prentice Hall.
11. Nair, M.C. & Balakrishnan, S. Beneficial fungi and their utilization. Scientific Publishers, Jodhpur.
12. Nash, T.H. Lichen Biology. Cambridge University Press.

13. Webster, John. Introduction to Fungi. Cambridge University Press.

Microbiology

1. Introduction - main groups of microorganisms and their characteristics - prions, viroids, viruses, bacteria, mycoplasmas and actinomycetes.
2. Bacteria - classification based on Bergey's Manual. Archaeobacteria and Eubacteria. Morphology, ultra-structure, nutrition, genetics
3. Plasmids and their characterization.
4. Cyanobacteria- salient features, morphology, ultrastructure, classification and economic importance.
5. Viruses- General account of plant and animal viruses, bacteriophages and their classification. Isolation, purification, infection, replication and transmission of plant viruses. Detailed study of TMV and T4 Phage.
6. Microbial ecology- microbiology of rhizosphere and phylloplane. Sewage disposal, bioremediation and water purification. Detection of microbes in air and water.
7. Agricultural microbiology - management of agricultural soils, biofertilizers, biopesticides.
8. Food Microbiology - Food spoilage and preservation methods. Microbiology of fermented food - dairy products, bread and other fermented plant products. Microorganisms as source of food- single cell protein.
9. Industrial Microbiology - Production of alcohol, vinegar, antibiotics, vitamins, steroids, vaccines, organic acids, amino acids.

References

1. Adams, M R & Moss, M.O. Food Microbiology. New Age International Publishing Ltd., New Delhi.
2. Brock, T. D. Biology of Microorganisms. Prentice Hall.
3. Campbell, R. Microbiology. ELBS-Edward Arnold, London.
4. Carpemter, P.L. Microbiology. W.B. Saunders & Company, Philadelphia.
5. Dubey, R.C. & Maheswari, D.K. A text book of Microbiology.
6. S. Chand. Desikachary. Cyanophyta- Monograph
7. Goodfellow, M. et.al. The Biology of Actinomycetes. Academic press.
8. Kumar, H.D. & Swati Kumar. Modern Concepts of Microbiology.
9. Mathew, R.E.F. Plant Virology, Academic press.
10. Pelozar, M.J., Chan, E.C.S. & Krieg, N.R. Microbiology. Tata Mc Graw Hill.
11. Sharma, P.D. Microbiology & Plant Pathology. Rastogi Publishers, Meerut.

Plant Pathology

1. Principles of Plant Pathology- Causal agents of plant diseases - Biotic causes (fungi, bacteria, virus, mycoplasma, nematodes, angiospermic parasites. Abiotic causes (nutrient and mineral deficiencies, effect of pollution). Koch's postulates. Latrogenic diseases. Seed pathology.

2. Details of different symptoms of plant diseases.
3. Process of infection- mechanical, physiological and enzymatic action. Penetration and entry of pathogens in to host tissue.
4. Host- parasite interaction. Enzymes and toxins in pathogenesis. Defense mechanisms in plants (structural and biochemical).
5. Details of different ways of spread and transmission of plant diseases- wind and water-mediated, seed borne and vector borne.
6. Plant disease management- exclusion, eradication and protection. Different pesticides and fungicides and their application. Biocides in plant protection.
7. Study of the following diseases with reference to the symptoms, causal organisms, disease cycle and control measures:
 Bunchy top of banana, Bacterial blight of paddy, Bud rot of coconut, Mahali of Arecanut, Powdery mildew of rubber, Abnormal leaf fall of rubber, tikka disease of Ground nut, Late blight of potato, Blister blight of tea, wheat rust, coffee rust, grey leaf spot of coconut, Phytophthora foot rot of pepper, rhizome rot of ginger and turmeric, angiospermic parasites-Viscum, Dendrophthoe.

References

1. Agrios, G.N. Plant pathology. 4th Ed., Academic Press.
2. Bilgrami, K.H. & Dube, H C. A Text Book of Modern Plant Pathology. Vikas Publishers, New Delhi.
 Chaube, H.S. & Ramji Singh. Introductory Plant Pathology. International Book Distributing Co., Lucknow.
3. Gareth-Jones, D. Plant Pathology: Principles and Practice. Open University Press.
4. Horsfall J.G. & Cowling E. B. (Ed.). Plant Disease: An Advanced Treatise. Academic Press.
5. Lucas, J. A. Plant Pathology and Plant pathogens. Blackwell.
6. Manners, J.G. Principles of Plant Pathology. Cambridge Univ Press.
7. Mehrotra, R.S. Plant Pathology. Tata Mc Graw Hill.
8. Pandey, B. P. Plant Pathology -pathogen and plant disease. S. Chand & Co.
9. Pathak, V.N., Khatri, N.K. & Pathak, M. Fundamentals of Plant Pathology. Agro-bios India.
10. Rangaswami, G. Diseases of Crop Plants of India. Prentice Hall India.
11. Tarr, S.A. J. The Principles of Plant Pathology. Winchester Press.
12. Wheeler, H. Plant Pathogenesis. Springer Verlag.
13. Wood, R.K.S. Physiological Plant Pathology. Blackwell

BOT1C03 – ANGIOSPERM ANATOMY, ANGIOSPERM EMBRYOLOGY, PALYNOLOGY & LAB TECHNIQUES

Contact Hours per Week: 6

Course Outline

Angiosperm Anatomy

1. Cell wall and its development. Chemistry of cell wall- cellulose, hemicellulose, polysaccharides, cell wall proteins, water. Organization of primary wall. Cytokinesis and growth. Plasmodesmata. Secondary wall chemical constituents- lignin, suberin, callose; organization of secondary wall.
2. Node - nodal patterns: Unilacunar, trilacunar, multilacunar and split lateral. Phylogenetic considerations. Leaf trace and branch trace- origin, departure; effect on stele and pith. Secondary growth in leaf traces.
3. Cambium: Development of vascular cambium and cork cambium in root and stem; cell types in vascular cambium, infected vascular cambia, seasonal variations in cambial activity; role of cambium in wound healing and grafting. Conversion of fusiform initials in to ray initials; cambium in arborescent monocotyledons (Liliflorae).
4. Development and differentiation: The structure of specialized cells. Vascular differentiation (procambium, residual meristem, interfascicular and intrafascicular cambia); acropetal and basipetal differentiation in leaves, stem and roots. Sieve tube differentiation. Control of phloem differentiation. Tracheary elements differentiation. Ultra-structure of phloem and xylem, brief account of transfer cells. Secondary wall thickening, cytoplasmic changes and autolysis. Control of differentiation. Genetic aspects- induction of vessel elements. induction of secondary xylem structure in relation to function in water conduction.
5. Anomalous secondary growth: Concepts; modification of the common type of vascular cambium, unequal activity of the vascular cambium. Successive cambia. Anomalous placement of vascular cambium. Discontinuous, unidirectional and bidirectional activity of cambium. Anomalous secondary growth in storage roots (Beet root, sweet potato).
6. Seedling anatomy: Concepts: anatomy of cotyledons, hypocotyl, seedling root, mesocotyl differentiation
7. Leaf anatomy: Unifacial, bifacial and centric leaf (onion); structure of epidermis, stomatal types; foriar sclerieds; oil cells; crystal idioblasts.
8. Anatomy in relation to taxonomy.
9. Wood anatomy- general account.

References

1. Easu, K. Plant Anatomy - Wiley Eastern Limited.
2. Fahn, A. Plant Anatomy. Pergamon Press.
3. Cutter, E.G. & Edward, E. Plant Anatomy: Experiment and Interpretations Part I and II.

4. Mauseth, J.D. Plant Anatomy - The Nenjamin Cumming Publishing Co.
5. Forester, A.S. Practical Plant Anatomy. D. Van Nostrand Company Inc.
6. Roberts, L.W. Cytodifferentiation in Plants - Cambridge University Press, Cambridge.

Angiosperm Embryology

1. Introduction to angiosperm embryology - structure of dithecous and monothealous anther.
2. Microsporogenesis: Structure and function of wall layers, role of tapetum in pollen development
3. Male gametophyte: Pollen mitosis, division of generative cells, heterospory.
4. Megasporogenesis: Megaspore triad, dyad, coenomegaspore.
5. Embryo sac- different types- ultra-structure of components- synergid and antipodal. Theories of the morphological nature of embryo sac
6. Pollination -Artificial pollination - ultra-structural and dis-ultrastructural and histo-chemical sigma. Significance of pollen - pistil interaction. Role of pollen wall proteins and stigma. In vitro pollination and fertilization.
7. Fertilization: Role of synergids - filiform apparatus, heterospermy and triple fusion.
8. Structure and development of typical dicot and monocot embryos- structure and function of suspensor.
9. Endosperm: classification and type- ruminant endosperm- mosaic endosperm- endosperm haustoria- physiology and cytology of endosperm.
10. Polyembryony - classification – practical value.
11. Apomixis - general account, genetics of apomixis.
12. Parthenocarpy -seedless fruits
13. Experimental embryology-embryo culture, anther culture, ovule culture.
14. Embryology in relation to taxonomy.

References

1. Bouman F. Ovule initiation, ovule development and seed coat a structure in angiosperms. Today and Tomorrow Publishers, New Delhi.
2. Bhojwani S.S. and Bhatnagar S.S. The embryology of Angiosperms. Vikas Publication, New Delhi.
3. Davis C.L. Systematic embryology of Angiosperms. John Wiley.
4. Eames A.J. Morphology of Angiosperms. Mc Graw Hill.
5. Johanson D. Plant Embryology. Waltham, Massachusetts.
6. John B.D. (Ed.). Embryology of Angiosperms. Springer Verlag.
7. Maheswari P. An introduction to the Embryology of Angiosperms. Mc Graw Hill.
8. Raghavan V. Experimental embryogenesis in plants. Academic Press.
9. Wardlaw C.W. Embryogenesis in Plants. Methusen, London.

Palynology

1. Introduction- contributions of Erdtman and P. K. K. Nair.
2. Development and structure of pollen wall. Pollen morphology and its application. Pollen evolution
3. Aero-palynology- methods of aerospore survey and analysis
4. Melittopalynology- nutritional and medical value of honey- unifloral and multifloral honey.
5. Recent advances in palynological studies- forensic-pollen allergy-oil exploration-paleopalynology.
6. Palynology in relation to taxonomy- eurypalynous and stenopalynous taxa.

References

1. Sripad N. Agashe. Palynology and its Application.
2. Kahinath Bhattacharya et. al. A Text Book of Palynology.

Laboratory Techniques

1. Study of the following instruments - their uses and principles:
 - a. Microscope: microscopic measurements - camera lucida, micrometry.
 - b. Microtomes- Sledge, Rocking, Rotary.
2. Killing, fixing and staining of plant tissues:
 - a. Important reagents and chemicals used in the preparation of fixatives and their properties.
 - b. Fixatives - FAA, Carnoy's fluid, chrome acetic, Nawaschins fluid, Craff, Flemings- composition, preparation and specific uses.
 - c. Dehydrating agents, clearing agents, mounting media. Examples and brief description.
 - d. Stains - classification, composition and specific uses - safranin, crystal violet, cotton blue, fast green, Orange - G, hematoxylin, carmine.
 - e. Brief account of vital staining.
 - f. Staining techniques - Double staining.
 - i. Saffranin - Fast green
 - ii. Crystal violet – Orange G
 - iii. Methods of embedding plant materials in paraffin wax - TBA method; embedding for Electron microscopy.
 - iv. Sectioning of embedded paraffin wax materials using Rotary Microtome.
 - v. Double staining of microtome serial sections embedding in paraffin wax - Saffranin - fast green; Crystal violet - Orange G / Erythrosin.
 - vi. Whole mounts - general account
 - vii. Maceration, smears
 - viii. Histochemical tests –
 - (1) PAS Test - insoluble polysaccharides. (2) Sudan black -lipids. (3) Fuelgen reaction - Nucleic

References

1. Peter Gray. Hand book of Basic microtechnique. Mcgraw – Hill.
2. John E. Sass. Botanical Microtechnique, Oxford & IBH Publishing Co.
3. John R. Baker. Principles of Biological Microtechnique –
4. A guide book to microscopical methods. A. V. Grimstone and R.J. Saker, Cambridge Univ. press.
5. K.V. Krishnamurthy. Methods in Plant Histochemistry.

BOT1L01 – PRACTICALS OF PHYCOLOGY, BRYOLOGY, PTERIDOLOGY, GYMNOSPERMS, MYCOLOGY & LICHENOLOGY

Contact Hours per Week: 3

Course Outline

Phycology

1. Collection, preparation and submission of algal herbarium (5 numbers).
2. Collection and study of the types mentioned below and their identification up to generic level using algal monographs:

Chlorophyta: Pediastrum, Scenidesmus, Hydrodictyon, Ulva, Cladophora, Pithophora, Bulbochaeta, Cephaleuros, Draparnaldiopsis, Bryopsis, Codium, Caulerpa, Halimeda, Desmids (Closterium, Cosmarium), Nitella.

Xanthophyta: Botrydium.

Bacillariophyta: Biddulphia, Coscinodiscus, Cymbella.

Phaeophyta: Ectocarpus, Dictyota, Padina, Turbinaria.

Rhodophyta: Batrachospermum, Gracilaria, Champia.

Bryology

1. Morphological and structural study of representative members of the following groups using whole mount preparations, dissections and transactions:

Asterella, Targionia, Cyathodium, Lunularia, Pallavicinia, Dumortiera, Porella, Anthoceros, Sphagnum and Bryum.

Pteridology

1. Collection, preparation and submission of five herbarium sheets of pteridophytes.
2. Study of vegetative and reproductive features of Lycopodium, Ophioglossum, Angiopteris, Osmunda, Lygodium, Ceratopteris, Pteris, Asplenium, Blechnum, Cyathea, Gleichenia, Trichomanes, Salvinia and Azolla.

3. Study of the following fossils: Rhynia, Lepidodendron, Sphenophyllum, Calamites, Calamostachys, Zygopteris and Anachoropteris.
4. Spore germination and development of prothallus in Knop's Agar medium.

Gymnosperms

1. Identification of petrifications, compressions, impressions: Lyginopteris, Heterangium, Medullosa, Trignocarpus, Glossopteris, Caytonia, Pentaxylon and Cordaites.
2. Study of vegetative and reproductive structures of Zamia, Ginkgo, Pinus, Cryptomeria, Cupressus, Araucaria, Agathis, Podocarpus, Cephalotaxus, Ephedra and Gnetum.

Mycology

1. Critical study of the following types with the help of fresh/preserved materials by making suitable micropreparations giving emphasis on systematic position, details of vegetative and reproductive structures: Stemonitis, Saprolegnia, Phytophthora, Albugo, Mucor, Pilobolus, Saccharomyces, Xylaria, Chaetomium, Peziza, Puccinia, Auricularia, Polyporus, Ganoderma, Lycoperdon, Dictyophora, Geastrum, Cyathus, Aspergillus, Curvularia, Alternaria, Fusarium, Colletotrichum, Parmelia, Usnea.

Practical records:

Submission of certified record of practicals at the time of terminal evaluation.

Field work:

2 days of field work for the in-situ study of the types of the above areas of study and submission of a field report.

BOT1L02 – PRACTICALS OF MICROBIOLOGY, PLANT PATHOLOGY, ANGIOSPERM TAXONOMY, ANGIOSPERM EMBRYOLOGY, PALYNOLOGY & LAB TECHNIQUES

Contact Hours per Week: 3

Course Outline

Microbiology

1. Test for the presence of coliform bacteria in contaminated water.
2. Isolation of Eubacteria and Cyanobacteria from soil by dilution plate method.
3. Isolation of pure bacterial culture by streak plate method.
4. Staining of bacteria (negative staining, Gram staining and spore staining).
5. Demonstration of bacterial motility by hanging drop method.

6. Morphological studies on Scytonema, Aphanocapsa, Spirulina, Oscillatoria, Anabaena.

Plant Pathology

1. Submission of five herbarium sheets of pathological specimens.

2. Detailed lab study of the following diseases:

Bunchy top of banana, Bacterial blight of paddy, Bud rot of coconut, Mahali of Arecanut, Powdery mildew of rubber, Abnormal leaf fall of rubber, tikka disease of Ground nut, Late blight of potato, Blister blight of tea, wheat rust, coffee rust, grey leaf spot of coconut, Phytophthora foot rot of pepper, rhizome rot of ginger and turmeric, angiospermic parasites- Viscum and Dendrothoe.

3. Technique of isolation and pure culture of pathogens.

Angiosperm Anatomy

1. Study of anomalous secondary growth in roots and stems of Aristolochia, Strychnos, Amaranthaceae, Nyctaginaceae, Bignoniaceae and Agavaceae.

2. Nodal anatomy of different types.

3. Leaf anatomy: epidermal peels and TS of lamina.

Embryology

1. Study of anther development of Datura.

2. Preparation of dissected whole mounts of microsporangium.

3. Study of megaspore mother cell, megaspore and embryo sac. 4. Study of the receptivity of stigma and in situ germination of pollen.

5. Dissection of stages in the development of embryo and endosperm.

6. Pollen germination using hanging drop technique.

7. Demonstration of intra ovarian pollination.

Palynology

1. Analysis of honey for microscopic examination of pollen.

2. Calculation of percentage of viable pollen by using T Z test. 3. Study of pollen wall by acetolysis.

Lab Techniques

1. Measurement of microscopic objects - Micrometry.

2. Camera lucida drawing - calculation of magnification

3. Double stained permanent sections - free hand section, Microtome serial sections.

4. Preparation of whole mounts, macerations and smears.

5. Submission of 10 permanent slides - which should include microtome serial sections, free hand sections, macerations, whole mounts and smears.

Practical records:

Submission of certified record of practicals at the time of terminal evaluation.

Field work:

2 days of field work for the in-situ study of the types of the above areas of study and submission of a field report.

SEMESTER II**BOT2C04 – CELL BIOLOGY, MOLECULAR BIOLOGY & BIOPHYSICS**

Contact Hours per Week: 6

Course Outline**Cell Biology**

1. The nucleus. Interphase nucleus- Chromatin organization- nucleosomes, scaffold. Organization of eukaryotic chromosome. Heterochromatin- constitutive, facultative and condensed. Euchromatin. Satellite DNA. Chromosome banding and its significance.
2. Cell reproduction: Cell cycle. Specific events G₁, S, G₂ and M phases. Significance of G₀. Control of cell cycle. Significance. Gene expression during cell cycle. Mitotic Inducers.
3. Meiosis: types, synaptonemal complex, significance of meiosis. Genetic control and consequences of meiosis. Restriction points and check points. Cell cycle regulation of meiotic events- behaviour of sex chromosomes in meiosis- suppression of DNA replication between Meiosis I and II. Meiotic defects and human diseases.
4. Programmed cell death- necessity, classes, signals. Genetic analysis of cell death. Proteins regulating apoptosis. Pathways leading to cell death- significance. Aging- cellular and extracellular. Cell signaling.
5. Cell interactions-communication, recognition and adhesion. Application.
6. Cellular differentiation and specialization. General characteristics, intrinsic interactions- Nucleo-cytoplasmic. Extrinsic interactions. Molecular mechanisms of cellular differentiations.
7. Cancer- carcinogenic agents. Phenotype of the transformed cell. Genetic basis of malignant transformation- oncogenes. Tumour suppressor genes. Cancer and cell cycle. Metastasis. Interaction of cancer cells with normal cells.

References

1. Cooper Jeffrey M. The Cell- A Molecular Approach. ASM, Washington.
2. Karp Gerald. Cell Biology. JohnWiley and Sons.

3. Derobertis. Cell and Molecular Biology.
4. Sadava R.
5. Pollard T.D. and Earn Shaw W.C. Cell Biology. Saunders.

Molecular Biology

1. Molecular biology of gene: Structure of DNA: Repetitive DNA; c-value paradox.
2. Replication of DNA: Enzymology of replication. Replication in prokaryotes and eukaryotes. Primosomes and replisomes. Telomerase and its function.
3. Gene expression: regulation of gene expression- Operon concept- Gene regulation in prokaryotes and eukaryotes- enhancers and silencers.
4. Protein synthesis: Transcription, post-transcriptional events. Infrons and their significance. Translation. Post translational events. Role of chaperons.
5. Mutation: Spontaneous and induced. Physical and chemical mutagens. Molecular mechanism of mutation. Mutation and cancer. Mutator and antimutator genes. DNA repairing mechanisms.
6. Molecular evolution: The origin of genomes. Evolution of new genes. Origin of eukaryotic genomes. Phylogenetics. Application of molecular phylogenetics.

References

1. Lewin Benjamin. Genes. Oxford University press.
2. Brown TA. Genomes. John Willey and Sons.
3. Snustad, Simmons and Jenkins. Principles of Genetics. John Willey and Sons.
4. Weaver and Hendrick. Genetics. Wm. C. Brown Publishers.
5. Hawkins J.D. Gene Structure and Expression. Cambridge University Press.

Biophysics

1. pH and buffer solutions- hydrogen ion concentrations and pH, dissociation of acids and bases. Measurement of pH using organic indicator molecule and potentiometric method. Functions of buffers in a biological system. Use of buffers in biological and biochemical research. pH and life. Henderson and Hasselbalch equation.
2. Chromatography: Principles of chromatography. Types of chromatography (Brief account).
3. Electrophoresis: Electrophoretic mobility, principles, PAGE, Agarose gel electrophoresis. Separation and detection of macromolecules by electrophoresis. Electrophoretic apparatus, technique and procedure.
4. Centrifugation - Theory of centrifugation. Centrifuge- Types, Methodology of centrifugation, applications.
5. Colorimetry and spectrophotometry: Beer-Lamberts law. Measurement of extinction. Calorimeters and spectrophotometers. Techniques and applications in biological and biochemical research. Comparison

between colorimetry and spectrophotometry.

6. Radiobiology: Autoradiography. principles, types. Methods and applications in biological research.
7. Immunochemistry: Immune response. Antigens- Antibodies. Histo-incompatibility antigens; Structure of IgG. Immunochemical assays - RIA, ELISA.
8. Cryobiology: Freeze drying (lyophilization) - applications.

References

1. Hoppe, W. (Ed.). Biophysics. Springer Verlag.
2. Rogers, A.W. Techniques of Autoradiography. Elsevier.
3. Roy, R.N. A Text Book of Biophysics. New Central Book Agency Pvt. Ltd, Calcutta.
4. Sasidharan, A. Selected Topics of Biophysics. Frontier Area Publishers.
5. Slayter, E.M. Optical methods in Biology. Wiley Intersciences.
6. Wong, C.H. Radiation Tracer Methodology in Biophysical Sciences. Prentice Hall.
7. Plummer, D. An introduction to Practical Biochemistry. Tata Mc Graw Hill, New Delhi.

BOT2C05 – CYTOGENETICS, GENETICS, BIOSTATISTICS, PLANT BREEDING & EVOLUTION

Contact Hours per Week: 6

Course Outline

Cytogenetics

1. Cytogenetics of aneuploids, euploids and structural heterozygotes: Effect of aneuploidy on phenotype. Transmission of monosomics and trisomics and their uses. Breeding behaviour and genetics of structural heterozygotes; translocation heterozygotes; Robertsonian translocation; B-A translocation. Karyotype-concepts and its importance. Structural chromosome aberrations- types and significance in evolution. Heteroploidy, aneuploidy, monosomy, trisomy (primary, secondary, tertiary and compensating). Nullisomy. Uses of aneuploidy in cytogenetics. Euploidy- autopolyploidy, allopolyploidy and segmental allopolyploidization. Role of aneuploidy and euploidy in evolution.
2. Molecular cytogenetics: Multigenic families and their evolution; in situ hybridization- concept. Computer assisted chromosome analysis, chromosome micro-dissection and micro-cloning; flow cytometry.
3. Polytene and lampbrush chromosomes- cytogenetic importance.
4. Supernumerary chromosomes: B-chromosomes.

References

1. Alberts B., D. Bray, J. Lewis, K. Roberts and J. D. Watson. Molecular Biology of the Cell Garland Publishing Inc. New York.

2. Atherly A.G., J.R. Girton and J.F. McDonald. The Science of Genetics. Saunders College Publishing, Fort Worth, USA.
3. Burnharm C.R. Discussions in Cytogenetics. Burgess Publishing Co., Minnesota.
4. De Robertis E.D.P. and De Robertis E.M.F. Cell and Molecular Biology ISBN, Hong Kong.
5. Dupraw E.J. DNA and Chromosomes. Holt, Rinehart and Winston Inc. New York.
6. Hart D.L and E.W. Jones. Genetics: Principles and Analysis. Jones & Bartlett publishers, Massachusetts, USA.
7. Khush, G.S. Cytogenetics of Aneuploids. Academic Press.
8. Karp G. Cell and Molecular Biology: Concepts and Experiments. John Wiley & Sons, Inc. USA.
9. Lewin B. Gene. Oxford University Press, New York, USA.
10. Lewis R. Human Genetics: Concepts and Applications. WCB Mc Graw Hill, USA.
11. Malacinski G.M and D. Freifelder. Essentials of Molecular Biology. Jones and Bartlett Publishers Inc., London
12. Rieger R., A. Michaelis and M. M. Green Glossary of Genetics and Cytogenetics - Classical and Molecular. Springer-Verlag, New York.
13. Swanson C.P., T. Merz, and J. W. Young. Cytogenetics. Prentice Hall.

Genetics

1. Relevance of Mendelism in modern genetics. A critical evaluation of Mendelism on the basis of modern concept of genes.
- 2 Linkage and gene mapping. Three- point test cross; linkage map; interference; tetrad analysis and centromere mapping. Linkage in humans. Pedigree analysis. Genetic recombination and mapping of genes in bacteria and bacteriophages.
3. Mobile genetic elements: Transposable elements in bacteria. IS elements. Tn elements. Cmp site transposon. Cepia and P elements in Drosophila. Ac, DS and Mu elements in maize. Retrotransposons- Molecular characteristics and significance in development and evolution.
4. Extranuclear inheritance: Analysis of mitochondrial and chloroplast genomes and their utility. Cytoplasmic male sterility.
5. Quantitative genetics: Polygenic inheritance, heritability and its measurements. QTL mapping.
6. Population genetics: Systems of mating. The Hardy-Weinberg principle. Estimation of gene frequencies. Factors affecting equilibrium: natural selection, mutation, migration and genetic drift.
7. Human genetics: Human pedigree analysis, Lod score for linkage testing. Karyotype; genetic disorders.

References

1. Snustad, Simmons and Jenkins. Principles of Genetics. John Willey and Sons.
2. Weaver and Hendrck. Genetics. Wm. C Brown Publishers.

3. Goodenough. Genetics. Saunders College Publishing.
4. Stansfield. Theory and Problems of Genetics. Mc Grow Hills.
5. Strickberger. Genetics. Macmillan.
6. Burnet L. Essential Genetics. Cambridge University Press.
7. Friefelder. Microbial Genetics. Narosa Publishing House.
8. Gardner, Simmons and Snustad. Principles of Genetics. John Wiley and Sons, New York, USA.
9. Singh B.D. Fundamental of Genetics. Kalyani Publishers, New Delhi.

Biostatistics

1. The science of statistics and its applications in biological research.
2. Types and collection of data- Census and sampling- theory and methods.
3. Tabulation and presentation of data- diagrammatic and graphic presentation.
4. Analysis of data- central tendencies.
5. Measures of dispersion - Range, quartile deviation, mean deviation, standard deviation and standard error.
Relative measures of dispersion - coefficient of variation.
6. Tests of significance- formulation and testing of hypothesis- testing the probability of committing type 1 and type 2 errors. z test, t test, chi-square test.
7. Analysis of variance- one way classification and two-way classification, F test, F value calculation, F table.
8. Correlation and Regression analysis- coefficient of correlation- significance testing. Rank correlation.
Lines of regression- coefficient of regression.
9. Experimental designs- designing an experiment- CRD, RBD, LSD. Factorial experiments.
10. Probability- application of the principles of probability- theorems of probability- applications- Probability distributions- binomial, multinomial, normal and poisson distributions.
11. Statistical softwares- SPSS, SPAR, MINITAB.

References

1. Chandal S.R.S. A Handbook of Agricultural Statistics. Achal Prakashan Mandir, Kanpur, India.
2. Das M.N. and N.C. Giri. Designs and Analysis of Experiments. Wiley Eastern Ltd.
3. Elhance and Elhance. Fundamentals of Mathematical Statistics. Kithab Mahal, New Delhi, India.
4. Gupta S.K and V.K. Kapoor. Fundamentals of Mathematical Statistics. Sultan Chand & Sons, New Delhi.
5. Gupta C.B. An Introduction to Statistical Methods. Vikas Publishing House Pvt. Ltd.
6. Kempthorne, O. An introduction to Genetic statistics. John Wiley and Sons Inc. New York.
7. Mather K. and J.L. Links. Biometrical Genetics. Chapman and Hall, London.
8. Panse, V.G and P. Sukatme. Statistical Methods for Agricultural Workers. ICAR, New Delhi.

9. Rao C.A. Advanced Statistical Methods in Biometrical Research. Wiley and Sons, New York.
10. Singh P. and S.S. Narayanan. Biometrical Techniques in Plant Breeding. Kalyani Publishers, New Delhi.
11. Singh R.K. and Chaudhary B.D. Biometrical Methods in Quantitative Genetic Analysis. Kalyani Publishers, New Delhi.
12. Daniel W.W. Biostatistics- A foundation for Analysis in Health Sciences.

Plant Breeding

1. Introduction and objectives.
2. Organizations involved in plant breeding.
3. Breeding systems in sexually propagated plants- Floral Biology and its significance in plant breeding. Sterility and incompatibility systems.
4. Genetic resources- centers of crop genetic diversity. In situ and ex situ conservation; cryopreservation of germplasm.
5. Conventional methods of plant breeding:
 - Domestication of wild plants- changes under domestication.
 - Plant introduction- history, types, principles, plant introduction agencies in India- rules and regulations. Major achievements
 - Selection- selection methods in sexually and vegetatively propagated species. Selection in segregating populations. Major achievements.
 - Hybridization- history, objectives, techniques, consequences and major achievements. Heterosis breeding- genetic basis of heterosis and inbreeding depression.
6. Modern methods of plant breeding:
 - Mutation breeding- history, methodology, applications, merits, demerits and achievements. Polyploidy breeding- methodology, applications, merits, demerits and achievements.
 - Biotechnological approaches in plant breeding- Molecular markers and their uses- Transgenic plants- critical evaluation.
7. Breeding for special purposes: Resistance breeding- a brief account of disease resistance, pest resistance, stress resistance- achievements. Quality breeding- objectives and achievements.
8. Biometrical techniques in Plant Breeding- analysis of variability, heritability, genetic advance and combining ability.
9. IPR- Protection of plant variety and farmers' right act.

References

1. Allard R.W. Principles of Plant Breeding. John Wiley and Sons, New Delhi.
2. Chahal G.S. and Gosal S.S. Principles and Procedure of Plant Breeding. Narosa Publishing House, New Delhi.

3. Jain H.K. and Kharkwal M.C. Plant Breeding- Mendelian to Molecular Approaches. Narosa Publishing House, New Delhi.
4. Roy D. Plant Breeding- Analysis and Exploitation of Variation. Narosa Publishing House.
5. Hayward M.D., Bosemark N.O. and Romagosa I. Plant Breeding- Principles and Prospects. Chapman & Hall.
6. Gupta S.K. Plant Breeding- Theory and Techniques. Agrobios (India), Jodhpur.
7. Khan M.A. Plant Breeding. Biotech Books, New Delhi.
8. Stoskopf N.C. Plant Breeding- Theory and Practice. Scientific Publishers (India), Jodhpur.
9. Sharma J.R. Principles and Practices of Plant Breeding. Tata Mc Graw Hill.
10. Chopra V.L. Breeding Field Crops. Oxford & IBH.
11. Mohanan K.V. Essentials of Plant Breeding. PHI Ltd., New Delhi.
12. Mohanan K.V. Essentials of Plantation Science. Penta Book Publishers, Calicut, Kerala.

Evolution

1. The concept of evolution- evidences of evolution- geological time scale and evolution.
2. Origin of life- theories and experimental evidences- chemical evolution and biological evolution.
3. Evidences of evolution.
4. Theories of evolution- Pre-Darwinian, Darwinian and Post Darwinian theories - Modern synthetic theory of evolution.
5. Reproductive isolation and the origin of species.
6. Evolution at the molecular level.

BOT2C06 –PLANT ECOLOGY, CONSERVATION BIOLOGY, PHYTOGEOGRAPHY & FOREST BOTANY

Contact Hours per Week: 6

Course Outline

Plant Ecology & Conservation Biology

1. Habitat Ecology: Salient features of terrestrial (Biomes), fresh water (Limnology), wet land and marine habitats.
2. Productivity and Energy flow: Concepts, limits and process of primary production; methods of productivity measurements: global trends in primary productivity, energy flow models.
3. Population characteristics: density, natality, mortality, distribution, biotic potential, carrying capacity, aggregation and dispersal, ecotone and edge effect.
4. The environment and its pollution- types (land, air and water). Effect on living organisms. Control with emphasis on biological methods. Environmental hazards.

5. Threats to the global environment- greenhouse effect, ozone depletion, El-Nino and La Nina effects.
6. Environment impact assessment (EIA) and assessment of environmental hazards- remote sensing.
7. Problems of conservation; causes of threat to environment- human interference, deforestation, habitat destruction, overexploitation of resources.
8. Identification of threatened plants; red list categories- extinct, endangered, vulnerable, rare and out of danger. Extinction process. Hot spots, keystone species and flagship species.
9. Strategies for conservation: in situ and ex situ conservation, biosphere reserve, national parks, wildlife sanctuaries. Gene banks, cryopreservation, seed banks.
10. Afforestation- social forestry, agroforestry. International biological programme (IBP), Man and biosphere programme (MAB), IUCN, world environment day, wild life preservation act (1972), Indian forest (conservation) act (1980) and United Nations Environment Programme. Environment Protection Acts.
11. Environmental awareness- role of government and NGOs. -Gaia hypothesis
12. Biodiversity- significance at Local, National and Global levels. Deep ecology (Paradigm shift from anthropocentric ecology to ecocentric ecology. National heritages.

References

1. Negi, S.S. Hand book of National Parks and Sanctuaries in India.
2. M.P. Nair and P.K Sastry - Red data book of Indian plants.
3. Mehrotra and B.K Suri - Remote sensing for environment and forest management.
4. Negi S.S - Biosphere reserves in India.
5. Lucas and Syngé - IUCN Red data book. IUCN, Stockholm
6. Dasman R.F - Environmental Conservation.
7. Odum E.P. Fundamentals of ecology
8. Odum E.P. Basic principles of ecology
9. Misra K.R. Ecology workbook.
10. Puri G.S. - Indian Forest Ecology Volumes I and II. Oxford & IBH.
11. Clarke G.L - Elements of Ecology.
12. Chhatwal G.L. Encyclopedia of environmental biology.
13. Ray P.K. - Pollution and Health. Willey-Eastern Ltd, New Delhi.
14. Michael P.- Ecological methods for field and laboratory investigations. Tata McGraw Hill, New Delhi.
15. Kershaw K.A. Quantitative and Dynamic Plant Ecology. ELBS.

Phytogeography

1. Patterns of plant distribution: continuous distribution: circumpolar, circumboreal, circum austral, pan tropical.
2. Discontinuous distribution: Theory of land bridges, theory of continental drift, theory of glaciation.

3. Endemic distribution (neoendemic, paleoendemic), age and area hypothesis.
4. Phytochoria of world and India.

References

1. Ronald Good. The geography of flowering plants. Lcngmans.
2. Bharucha F.R. A text book of plant geography of India. Oxford University Press.
3. Puri G.S. Indian Forest Ecology, Vol I, II. Oxford, New-Delhi.

Forest Botany

1. Forest- Definitions. Study of various types of forests in the world and in India.
2. Forest products- Major and minor with special reference to Kerala.
3. Influence of forests on environment. Consequence of deforestation and industrialization- sustainable utilization of bioresources.

References

1. Agarwal A. P. Forests in India. Oxford & IBH.
2. Gregorv G. R. Forest products, production, trade and consumption, quantity and value of raw materials requirements. Ford foundation, New-Delhi.
3. Puri G.S. Indian Forest Ecology Vol. I& II. Oxford & IBH.
4. Champion G. H. and Seth S.K. A revised survey of the forest types of India.

BOT2L03 – PRACTICALS OF CELL BIOLOGY, MOLECULAR BIOLOGY, BIOPHYSICS, CYTOGENETICS

Contact Hours per Week: 3

Course Outline

Cell Biology

1. Study of Mitosis in root tip cells.
2. Pre-treatment of root tips with colchicine /hydroxy quinoline /paradichlorobenzene and study of chromosomes in Chlorophytum, / Zea mays/ Crotalaria/ Cyanotis.
3. Isolation of plastids and mitochondria.
4. Chromosome banding

Molecular Biology

1. Working out problems from molecular genetics.
2. Isolation of nucleic acid and identification of histones by SDS-PAGE.
3. Isolation of plant DNA and its quantification by spectrophotometric/ calorimetric method.

4. Immunological techniques: ELISA and Western Blot.

Biophysics

1. Preparation of buffers and measurement of pH using pH meter.
2. Determination of isoelectric pH.
3. Paper chromatography: Separation of sugars.
4. Thin layer chromatography- separation of amino acid mixtures.
5. Calorimetric and spectrophotometric estimation of proteins by Biuret / Lowry's method.
6. Estimation of amino acid by ninhydrin method (colorimetric).

Cytogenetics

1. Induction of polyploidy using colchicine; different methods of the application of colchicine.
2. Effect of induced and spontaneous polyploidy on plant phenotype, meiosis, pollen and seed fertility and fruit set.
3. Preparation of karyotype and ideogram of plant meristematic cells.
4. Cytological studies in callus tissues.
5. Study of meiosis in translocation heterozygotes (Rheo discolor)
6. Study of polytene chromosomes.

Preparation of lab record and submission for valuation.

Visit to a reputed molecular biology lab and submission of a report.

BOT2L04 – GENETICS, BIOSTATISTICS, PLANT BREEDING, PLANT ECOLOGY, CONSERVATION BIOLOGY, PHYTOGEOGRAPHY & FORET BOTANY

Contact Hours per Week: 3

Course Outline

Genetics

1. Problems from linkage, tetrad analysis, quantitative genetics and population genetics.

Biostatistics

1. Problems from Mean, standard deviation, Coefficient of variation, tests of significance and correlation analysis.
2. Use of computer programmes for statistical analysis.

Plant Breeding

1. Study of floral morphology and flower structure in crop plants- rice, cashew, pulses, Solanum, Capsicum.
2. Practice of hybridization technique in self- and cross-pollinated plants mentioned in (1).
3. Biometrical techniques in Plant Breeding- analysis of variability.

Ecology and Conservation biology

1. Determination of food chains and food web in aquatic ecosystem.
2. Determination of the minimum size of the quadrat suitable for an area using species area curve method.
3. Determination of the Importance Value Index (IVI) of plant species in the community by quadrat, line and belt transect methods.
4. Comparative study of polluted and non-polluted aquatic ecosystems.
5. Visit to a meteorological station, national park or wild life sanctuary, sewage treatment unit and major construction site.
6. Estimation of dissolved oxygen content in the water sample by Winkler's method.
7. Determination of primary production in water samples by light and dark bottle method (Winkler's method).
8. Determination of dissolved carbon dioxide content in water samples.
9. Determination of frequency of plant species of an area and heterogeneity of vegetation using transect method.

Phytogeography

1. Identification of the various floristic and vegetational regions of the world and India in maps.

Forest Botany

1. Study of the major and minor forest products of Kerala and their uses.

Preparation and submission of lab record

Visit to one plant breeding station and one ecologically sensitive area and submission of reports

SEMESTER III

BOT3C07 – PLANT PHYSIOLOGY, METABOLISM & BIOCHEMISTRY

Contact Hours per Week: 6

Course Outline

Plant Physiology

1. Water and plant cells: Properties of water, hydrogen bonding, polarity, cohesion and adhesion. The concept of water potential. Water movements in cells and tissues. Soil-plant atmosphere continuum. Transpiration, stomatal movement, modern theories of stomatal mechanism. The ascent of xylem water and the uptake of water by roots. Absorption of mineral ions- solute absorption.
2. Plants and nitrogen: The nitrogen cycle. Biological nitrogen fixation, symbiotic nitrogen fixation in leguminous plants. Biochemistry of nitrogen fixation. Export of fixed nitrogen from nodules. Genetics of nitrogen fixation. Nitrogen assimilation, assimilation of nitrate. Nitrogen nutrition -agricultural and ecological aspects. Biosynthesis of amino acids- reductive amination and transamination. GDH and GS/GOGAT pathway.
3. Photosynthesis: Absorption and fate of light energy, absorption and action spectra. Photoreceptors- chlorophylls, carotenoids, phycobilins. Bioenergetics and the light dependent reactions of photosynthesis. Photosynthetic electron transport and photophosphorylation. The two pigment systems, Z-scheme, water oxidizing clock. The photosynthetic carbon reduction cycle, C3, C2, C4 and CAM metabolism and ecological significance.
4. Translocation and distribution of photo assimilates. Phloem transport, Sources and sinks, mechanism of translocation. Phloem loading and unloading, distribution of assimilates. Translocation of xenobiotic chemicals.
5. Patterns in plant development: Growth, differentiation, and development. Genetic control and hormonal regulation of development. Seed germination- physiology of hormones in plant development- auxins, gibberellins, cytokinins, abscisic acid and ethylene. Role of vitamins and nutrients.
6. Photomorphogenesis: Phytochrome: chemistry and physiological effects. Mechanism of phytochrome and gene action. Cryptochromes and blue light effect.
7. Stress Physiology: Types of stress- water, temperature, salt, stresses caused by pests and pathogens and pollutants.

References

1. William G. Hopkins. Introduction to Plant Physiology. John Wiley & Sons Inc.
2. Lincoln Taiz and Eduardo Zeiger. Benjamin/Cumming Publishing Company Inc. New York.

Metabolism

1. Enzymes: General aspect, classification, Michaelis - Menton equation and its significance. Mechanism of enzyme action, co-enzymes, inhibition, regulation, allosteric enzymes, covalently modulated enzymes. Kinetics of enzyme catalysis. Isoenzymes.
2. Intermediary metabolism: Anabolism, catabolism, amphibolic pathways and anapleurotic reactions. Link

between primary metabolism and secondary metabolism. Bioenergetics and thermodynamics.

3. Catabolism of hexoses: Glycolysis- two phases, overall balance sheet, regulation; fate of pyruvate under aerobic and anaerobic conditions. Pentose phosphate pathway-multifunctional pathway (significance). Tricarboxylic acid cycle: Formation of acetate, reaction of citric acid cycle, anapleurotic reactions of citric acid cycle. Regulation of citric acid cycle. Glyoxylate cycle. Amphibolic nature of TCA cycle.
4. Oxidation of fatty acids. Activation and entry of fatty acids, Beta oxidation of saturated and unsaturated fatty acids. Regulation.
5. Oxidation of amino acids and entry to TCA cycle.
6. Oxidative phosphorylation: Electron transfer reactions in mitochondria. Electron carriers, multienzyme complexes, ATP synthesis. Regulation of oxidative phosphorylation. Shuttle systems- Alternate pathways -Thermogenesis.
7. Carbohydrate biosynthesis: Gluconeogenesis, biosynthesis of starch, glucose and other carbohydrates. Involvement of NDP- sugars. Regulation.
8. Lipid biosynthesis: Biosynthesis of fatty acids. Triacylglycerols, phospholipids and isoprenoids. Regulation.
9. Biosynthesis of nucleotides: PRPP and its significance. Purine and pyrimidine biosynthesis. Precursors and regulation. Conversion of NMP to NTP. Biosynthesis of deoxyribonucleotides.
10. Secondary metabolism: Main pathways and their relation to primary-metabolism.

References

1. Lehninger. Principles of Biochemistry, Macmillan, U.K.
2. Geoffrey Zubay. Biochemistry. Macmillan Publishing Company, New York.
3. Trevor Palmer. Enzymes- Biochemistry, Biotechnology and Clinical Chemistry. Norwood Publishing, Chichester.

Biochemistry

1. The molecular logic of life.
2. The chemical unity of diverse living organisms.
3. Weak interactions in aqueous systems and the fitness of the aqueous environment for living organisms. 4. Biomolecules: a- Carbohydrates- Classification, structure and functions of simple sugars and compound carbohydrates. Sugar derivatives of biological importance. b- Lipids. Classification- storage and structural lipids; lipids in membranes; the supramolecular architecture of membranes. c- Amino acids, peptides and proteins. Amino acids: classification based on polarity; properties. Covalent structure of proteins. Three-dimensional structure of proteins. Protein- tertiary and quaternary structures. Denaturation and renaturation. Functions of protein. d- Nucleotides and nucleic acids. Chemistry- structure of nucleotides-

Other functions of nucleotides.

5. Secondary metabolites: Secondary metabolites, their physiological roles. Significance- ecological and phylogenetic importance.

References

1. Lehninger. Principles of Biochemistry, Macmillon, U.K.
2. Geoffrey Zubay. Biochemistry. Macmillen Publishing company, Newyork.
3. Sadasivam and Manickam. Biochemical Methods. New Age International Publishers. New Delhi.
4. David T. Plummer, An Introduction to Practical Biochemistry. Tata McGrow Hill.

BOT3C08 – ANGIOSPERM MORPHOLOGY, ANGIOSPERM TAXONOMY AND PLANT RESOURCES

Contact Hours per Week: 6

Course Outline

Angiosperm Morphology

1. A critical study of the current ideas on the origin of Angiosperms with special reference to their ancestral stock, time and place of origin.
2. The concept of primitive angiosperm flower. Origin and evolution of flower, co-evolution of flowers vis-a-vis pollinators.
3. Origin and evolution of structure and morphology of stamens, nectarines and nectar.
4. Origin and evolution of carpels: different types- concept of foliar origin of carpels; types of ovary; evolution of placentation types- inferior ovary- foliar and axial concepts.
5. Role of floral anatomy in interpreting the origin and evolution of flower and floral parts

References

1. Eames, E.J. Morphology of Angiosperms. Mc Graw Hills Book Co. New York.
2. Bamard, C. The interpretation of Angiosperm flower. Aust. J. Sci.24:64-72. (1961).
3. Manilal, K.S. Vascularization of corolla in Compositae. J.Indian Bot. Soc.59: 189-196
4. Meeuse, A.D. J. Some fundamental principles of interpretive floral morphology. Lntemational Sci. Publ. Hissar.
5. Melville, R. New theory of Angiosperm flower. Nature 188: 14-18. (1960).
6. Puri, V. Inferior ovary. Phytomorpholgy2:122. (1952).
7. Sporne, K.R. The Morphology of Angiosperms. Hutchinsons Uni. Press, London.

Angiosperm Taxonomy

1. Principles of Taxonomy- Scope and importance of Taxonomy; systems of classification- artificial, natural and phylogenetic systems; phenetic versus phylogenetic systems; cladistics in taxonomy; APG system of classification.
2. Conceptual basis of classification- essentialism, nominalism, empiricism, phenetics and cladistics. phylogenetic and alternative; concept of genus; concept of family; infraspecific categories.
3. Definitions and terms: primitive and advanced; homology and analogy; parallelism and convergence; monophyly and polyphyly.
4. Taxonomic hierarchy- concept of taxa- species, genus and family- infra specific categories.
5. Plant nomenclature: history of nomenclature; polynomial and binomial systems; detailed study of salient features and major provisions of the International Code of Botanical Nomenclature. Effective and valid publication, rank of taxa, rule of priority and its limitations, typification, author citation, rejection of names and names of hybrids. A brief account of International Code of Nomenclature of Cultivated Plants.
6. Concepts of character: definition, classification of characters- analytical and synthetic, qualitative and quantitative; unit and multiple, good and bad; correlation of characters; character weighting.
7. Modern trends in Taxonomy: cytotaxonomy, chemotaxonomy, biosystematics and numerical taxonomy. Molecular taxonomy- DNA bar coding in plants.
8. History and development of taxonomy in India. Classification of taxonomic literature- general indices, floras, icons, monographs, reviews-and journals; Herbarium- definition, steps involved in the development of herbarium- utility of herbarium and its maintenance- general account of regional and national herbaria with special reference to central National herbarium, Calcutta (CAL) and Madras Herbarium (MH). Botanical survey of India; Botanical gardens- types of gardens and importance of gardens in taxonomic studies- important national and international Botanical Gardens- Royal Botanical Garden, Kew; Indian Botanical Garden, Calcutta, National Botanical Garden, Lucknow, Tropical Botanic Garden, Trivandrum.

References

1. Cronquist A. Evolution and classification of flowering plants. Thomas and Nelson Co.
2. Cronquist. A. An integrated system of classification of flowering plants. New York.
3. Graf A.B. Tropica. Roehrs Company Publ. NJ, USA.
4. Harborne J.B. & Turner B.L. Plant chemosystematics. A.P., London.
5. Haywood W.H. & Moore D.M. Current concepts in plant taxonomy.
6. Rendle A.E. Classification of flowering plants.
7. Lawrance. G.H.M. Taxonomy of vascular plants. Oxford and IBH.

8. Sneeth P.H.A. umerical taxonomy. W.H. Freeman Co., São Francisco.
9. Sporne. K.R. The Morphology of Angiosperms. Hutchinson University Press, London.
10. Sivarajan V.V. Introduction to principles of plant taxonomy. Oxford and IBH.
11. Smith P.M. The Chemotaxonomy of plants. Edward Arnold, London.
12. Stace, C.A. Plant Taxonomy and Biosistematics. Edward Arnold, London.
13. Takhtajan, A. L. Diversity and classification of flowering plants. Columbia University Press, New York.
14. Woodland, D.W. Contempotary plant systematics. Prentice Hal, New Jersy.
15. Simpson M.G. Plant Systematics. Elsevier, Amsterdam.
16. Stebbins, G.L. Flowering Plants- Evolution above species level. Edward Arnord, London.

Plant Resources

1. A study of history, occurrence, morphology of useful part and overall chemical composition of the following:
 - a. Cereals & millets: rice, wheat, maize, sorghum, finger millet, pearl millet.
 - b. Pulses: Bengal gram, cluster bean, common bean, horse gram, cow pea.
 - c. Sugar yielding plants: sugar cane, beet root.
 - d. Starch yielding tubers: potato, tapioca, arrow root, yam, taro.
 - e. Fats & Oils: ground nut, coconut, castor, gingelly, mustard, oil palm.
 - f. Beverages: tea, coffee, cocoa.
 - g. Spices and Condiments: pepper, ginger, turmeric, coriander, cumin, fennel, fenu-greek, cardamom, nutmeg, cloves, cinnamon.
 - h. Fibre yielding plants: cotton, jute, coir.
 - i. Rubber yielding plants: pararubber.
 - j. Timber yielding plants: teak, rose wood, Artocarpus, Ailanthos, Xylia.
2. A study of the following medicinãl plants with reference to the chemical and pharmacognosic properties: neem, turmeric, Adhatoda, Rauwolfia, Catharanthes, Bacopa, nux-vomica, sweet flag, Saraca, wood apple, Indian myrobalans, liquorize.

References

1. Arora R.K. & Nayar, E. K. Wild relatives of crop plants in India. NBPGR Sci. Monograph No.7.
2. Bole, P.V. & Vaghani, Y. Field guide to common Indian trees. Oxford Uni. Press.
3. Chandel, K.P.S., Shukla, G. & Sharma, N. Biodiversity in medicinal and aromatic plants in India- conservãtion and utilization. NBPGR. New Deihi.
4. Chripeels, M.J. & Sadava, D. Plants, food and people. W..Freeman & Co. San Francisco.
5. ConwqyG. The doubly green revoluuiion: food for all in the 21st century. Penguin Books.
6. CS1R. The useful plants óf India. Publication and Information directorate, CSIR, New Delhi.

7. Kochar S.L. Economic Botany of the Tropics. Macmillon India Ltd.
8. Nair M.N.B. et al. (eds.) Sustainable management of non-wood forest products. Faculty of Forestry, Uni. Putra, Malaysia.
9. Padora R.S. and Arora R.K. Plant genetic resources and management. IPGRI Publication, South Asia office, NBPGR, Pusa Campus, New Delhi.
10. Indian Science Academy. Plant wealth in India. Special issue of proceedings, 1997.
11. Sahni, K.C. The Book of Indian Trees. Oxford Uni. Press, Mumbai.
12. Sharma, O.P. Hill's Economic Botany. Tata Mc Graw Hill Co., New Delhi.
13. Swaminathan M.S. & Kochar, S.L. (eds.). Plants and society. Macmillan Publication, London.
14. Thakur, R.S., Puri, H.S. & Husain, A. Major medicinal plants of India. Central Institute of Medicinal and Aromatic Plants, CSIR, Lucknow.

BOT3C09 – BIOTECHNOLOGY AND BIOINFORMATICS

Contact Hours per Week: 6

Course Outline

Biotechnology

A. Plant Tissue Culture

1. Basic concepts and history.
2. General account of laboratory facilities and management.
3. Media for in vitro culture, composition and their preparation.
4. Callus culture- selection of explants and medium- types of callus- growth profile of callus.
5. Cell culture - isolation of single cells- mechanical and enzymatic methods- measurement of growth of cells in suspension culture- viability tests.
6. Large scale cultivation of cells using bioreactors for secondary metabolite production.
7. Organogenesis- direct and indirect- factors affecting organogenesis.
8. Organ culture – apical/ axillary meristems, embryo, ovary, ovule, endosperm, anther, pollen and root cultures.
9. Applications of plant tissue culture - clonal propagation, somaclones, somatic hybrids, synthetic seeds, secondary metabolites, germplasm conservation –cryopreservation.

B. Genetic Engineering

1. Molecular analysis of gene and gene products: southern, northern and western blots- restriction maps- RAPD and RFLP. Chromosome walking and jumping. FISH. PCR and its applications. DNA fingerprinting. DNA chips.
2. DNA sequencing: Enzymatic methods. Gilbert and Maxam method. Messing's shot gun method.

Fluorescent detection and automation. The Human Genome Project.

3. Recombinant DNA Technology- Enzymes, vectors, gene-cloning strategies, construction and screening of gene and cDNA Libraries. Expression of cloned genes in bacteria and mammalian cells. Prospects and achievements.
4. Transgenic plants. Gene cloning strategies in plants. Vector dependent and vector independent methods. Identification and selection of transformed plants; the reporter enzyme technology.
Objectives and achievements- engineering for secondary metabolites; resistance against herbicides, pests, pathogens, stress - improved nutritional and status changes in plants. Plants as bioreactors; phytopolymers and biodegradable plastics; antisense RNA technology; transgene inactivation. Terminator and traitor technologies.
5. Cloning: objectives. Creation of transgenic animals- other developments in cloning. Human cloning. Ethics of cloning.
6. Patenting of genes and GMOs. Gene piracy. Ethics and biosafety aspects, rec DNA safety; IPR, biosafety protocols.

References

1. Walker J.M. and R. Rapley. Molecular Biology and Biotechnology: Panima Publishing Corporation.
2. Bernard R. Glick and Jack J. Pasternack. Molecular Biotechnology Principles and Applications of Recombinant DNA: ASM Press Washington
3. Brown T.A. Gene Cloning and DNA Analysis Blackwell Science Pub:
4. Primrose S.B. Molecular Biotechnology. Panima Publishing Corporation.
5. Maarten J. Chrispeels and D.E.Sadava. Plants, Genes and Agriculture. Jones and Bartlett Publishers.
6. Robert de la Pemere and Franck Seuret. Brave New Seeds: The threat of GM crops to farmers. Global Issues Series.

Bioinformatics

A. Computer application.

1. Computer in Science with special reference to biology, the scope and prospects.
2. Information super highway (Internet)- Information networks: Internet, World Wide Web. Web browsers, HTTP, HTML and URLs. Biological networks.
3. Online publications with special reference to biology, -electronic journals, books, downloading and uploading.)- Open Archive Initiative (www.openarchives.org), biomedcentral, pubmedcentral, freedom of scientific information access, e-access debate- concepts and implications, Free Software Movement, Free Software Foundation, GNU/Linux, etc. Online archives, databases, the Public Library of Science (www.publiclibraryofscience.org).

References

1. Online resources freely available at internet sites such as www.publiclibraryofscience.org
2. www.openarchives.org
3. www.pubmedcentral.gov
4. www.biomedcentral.com
5. www.nature.com/nature/debates/e-ccess/index.html

B. Bioinformatics

1. Introduction: Importance and scope.
2. Biological Databases
 - a. Nucleic acid databases: EMBL, GenBank- structure of GenBank entries. Specialized genomic resources, UniGene.
 - b. Protein sequence databases: PIR, SWISS-PROT, TrEMBL. Composite protein databases: NRDB, OWL. Secondary databases: PROSITE, PRINTS, BLOCKS, IDENTIFY. Structure classification databases- SCOP, CATH.
3. Database searching
 - a. Sequence database searching. EST searches. Different approaches to EST analysis. Merck/IMAGE, Incyte, TIGR. EST analytical tools. Sequence similarity, sequence assembly and sequence clustering.
 - b. Pair wise alignment technique: Comparison of sequences and sub-sequences. identity and similarity. Substitution matrices, BLOSUM, DOTPLOT and BLAST.
 - c. Multiple alignment technique: Objective, Manual, simultaneous and progressive methods. Databases of multiple alignments. PSI-BLAST, CLUSTAL-W.
4. Protein structure Prediction:
 - a. Secondary structure prediction. Chou- Fasman, J Pred.
 - b. Tertiary structure prediction: Comparative modelling - Modeller, RasMol.
5. Emerging areas of Bioinformatics: DNA Microarrays, functional genomics, comparative genomics, pharmacogenomics, chemoinformatics, Medical informatics.

References

1. Attwood T.K. and D.J. Argy-Smith. Introduction to Bioinformatics. Pearson Education.
2. Sundararajan S. and R. Balaji, Introduction to Bioinformatics. Himalaya Publishing House.

BOT3L05 – PRACTICALS OF PLANT PHYSIOLOGY, METABOLISM, BIOCHEMISTRY, ANGIOSPERM MORPHOLOGY, AND TAXONOMY

Contact Hours per Week: 3

Course Outline

Plant Physiology

1. Determination of water potential by tissue weight change method.
2. Extraction of leaf pigments and preparation of absorption spectra of chlorophylls and carotenoids.
3. Demonstration of Hill reaction.
4. Separation of leaf pigments by paper chromatography and column chromatography.
5. Effects of light intensity on photosynthesis by Wilmot's bubbler.
6. Determination of sugars and amino acids in germinating seed by TLC.
7. 7.Extraction of seed proteins based on solubility.
8. Biochemical analyses of leakages from seeds during germination.
9. Analyses of proline in water stressed plants.
10. Testing of seed viability by NBT test.
11. Changes in the reserve proteins during germination.

Metabolism

1. Extraction of enzyme: Any enzyme.
2. Effect of substrate on enzyme and determination of its K_m value.
3. pH dependent activity profile of enzymes.
4. Ammonium sulphate precipitation of enzymes.
5. Desalting of proteins by gel filtration using Sephadex G25/ dialysis
6. Separation of isoenzymes by native PAGE.
7. Determination of enzyme / protein sub units by SDS PAGE.
8. Metabolism of germinating seeds - changes in metabolisable carbohydrates.

Biochemistry

1. Qualitative tests for monosaccharides, reducing and non-reducing oligosaccharides, starch, amino acids and protein.
2. Quantitative estimation of reducing sugars and starch.
3. Qualitative tests for lipids. Emulsification, saponification, acrolein test, Boundouin's test.
4. Quantitative estimation of amino acids.
5. Quantitative estimation of protein by Biuret / Branford's /Lowry et al method.

6. Quantitative estimation of DNA and RNA (colorimetric / spectrophotometric)

7. Quantitative estimation of total phenolics.

Angiosperm Morphology

1. Preparation of cleared whole mounts of floral parts to show vasculature.

2. Examination of the following with the help of dissections and hand sections: Transmitting tissues/canals in style and stigma; Different types of ovaries; Different types of placentation, vasculature of androecium and gynoecium in special types of flowers.

Angiosperm Taxonomy

1. Familiarization with local flora and construction of keys – use of floras in identification up to species.

2. Study of diagnostic features of the families studied in the theory paper with special reference to their economic aspects.

3. Study of the following families with special reference to morphology of modified parts, economic importance, interrelationships and evolutionary trends: Magnoliaceae, Ranunculaceae, Menispermaceae, Nymphaeace, Polygalaceae, Caryophyllaceae, Clusiaceae, Sterculiaceae, Meliaceae, Sapindaceae, Rosaceae, Melastomaceae, Rhizophoraceae, Aizoaceae, Rubiaceae, Sapotaceae, Gentianaceae, Boraginaceae, Convolvulaceae, Scrophulariaceae, Pedaliaceae, Verbenaceae, Nyctaginaceae, Euphorbiaceae, Urticaceae, Casuarinaceae, Orchidaceae, Zingiberaceae, Amaryllidaceae, Commelinaceae, Araceae, Cyperaceae and Poaceae.

4. Dissection of at least two members of each family in the laboratory, making suitable sketches, describing them in technical terms and identifying them constructing appropriate floral diagrams.

5. Field study of three days under the guidance and supervision of teachers at an ecologically different locality and submission of a field study report certified by the teacher concerned. The report should contain ecology of flora of the area studied.

6. Collection of plant specimens following the standard means of plant collection for preparation of herbarium. Each student shall submit a minimum of 25 such herbarium specimens with QR code along with the field book for the Practical examination.

7. Problems in Bar Coding

BOT3L06 – PRACITCALs OF PLANT RESOURCES, BIOTECHNOLOGY AND BIOINFORMATICS

Course Outline

Plant Resources

1. Morphological study of the source plants mentioned in the theory syllabus and identification of the plants and plant products.

Biotechnology- A. Tissue Culture.

1. Preparation and sterilization of culture media.
2. Culturing of Carrot /Tobacco/Datura.
3. Estimation of cell growth in callus culture by fresh wt. and dry wt.
4. Induction of multiple shoots using axillary and apical meristems as explants.
5. Plantlet regeneration from callus.
6. Identification of secondary metabolites in cultures.

Biotechnology- B. Genetic Engineering

Isolation of DNA.

Bioinformatics

A. Computer Application

1. Acquiring basic computer operation and internet browsing skills in Windows and Linux platforms.
2. Acquiring basic word processing/ data entry skills using popular (both commercial and open source) packages such as MS-Word, K-Word, Open Word, PageMaker.
3. Acquire graphic processing skills using popular packages such as Photoshop, Corel Draw, Chem Draw.
4. Preparation of scientific presentations using packages such as MS-PowerPoint.
5. Use of statistical packages such as SPSS, Biostat, Origin, MS-Excel.

B. Bioinformatics

1. Acquisition of basic skills in Internet browsing
2. Use of web browsers and search engines.
3. Use of biological and bioinformatic websites Agris, Agricola, BIOSI S, CABWeb.
4. Visit to Bioinformatics websites: NCBI, SWISS PROT, PIR, PDB.

Submission of lab record

Submission of 10 plant products directly collected by the student from the field with a note on the source plant and plant part.

SEMESTER IV

**BOT4E01: To be selected by the Board of studies from the list appended as
Elective I**

**BOT4E02: To be selected by the Board of studies from the list appended as
Elective II**

L07: Practicals of Electives.

List of Electives for M. Sc. Botany CBCSS Programme:

A. Elective I

1. Advanced Angiosperm Taxonomy
2. Environmental Biology and Biodiversity Conservation
3. Plant Tissue Culture
4. Plant Physiology
5. Plant Cell and Molecular Biology
6. Genetics and Crop Improvement

B. Elective II

1. Molecular Plant Taxonomy
2. Pathology of Plantation crops and Spices
3. Genetic Engineering
4. Genomics and Proteomics
5. Genetic Engineering and Bioinformatics
6. Biotechnology in Crop Improvement

The board of studies selected the following Electives

BOT4E01 – ENVIRONMENTAL BIOLOGY AND BIODIVERSITY CONSERVATION

Contact Hours per Week: 6

Course Outline

1. **Population ecology:** Properties (concepts of rate, intrinsic rate of natural increase, carrying capacity, population fluctuations and cyclic oscillations, density independent and density dependent mechanisms of

population regulation, patterns of dispersion, Allee principle of aggregation and refuging, home range and territoriality, energy partitioning and optimization, r and K selection.

2. **Community ecology:** Types of interaction between two species, coevolution, evolution of cooperation, group selection, interspecific competition and coexistence, positive and negative interactions, concepts of habitat, ecological niche and guild.
3. **Human population:** Expansion and its causes, rich and poor nations, consequences, dynamics, Cairo conference 1994.
4. **Major global environmental challenges:** Acid rain, Ozone depletion, climate disruption, deforestation, land degradation and desertification, freshwater degradation and shortage, marine fisheries decline, loss of biological diversity and excess nitrogen.
5. **Global initiatives:** Stockholm conference (1972), Rio (1992), Ramsar convention (1971), Kyoto (1997), Johannesburg (2002), Stockholm (2011).
6. **Environmental Law- International and National:** The Environment Protection Act & Rules 1986; Water (Prevention & Control of Pollution) Act 1974; Biodiversity Act(2002).
7. **Thoughts on ecology:** Contributions of Buddha, Rabindranatha Tagore, Mahatma Gandhi, Rachel Carson, Gro Herlem Brundtland, Vandana Siva, Edward O Wilson, Aldo Leopald.
8. **Biodiversity:** a). Genetic diversity, agrobiodiversity and cultivated taxa, causes of decline, value of wild species, conservation practices- traditional (*upavana vinoda*, sacred groves, *sthalavrikshas*) and modern (*in situ* and *ex situ*). b). Biodiversity information management and communication- libraries, databases (taxonomic database working groups for plant sciences, data bases on biodiversity); distribution of biodiversity information, metadatabases, virtual libraries.
9. **Ecosystem capital- use and restoration:** Global perspective on biological systems; conservation, preservation and restoration. Biomes and ecosystems under pressure (forest biomes, ocean ecosystems).
10. **Habitat studies:** Wetlands (Ramsar sites), mangroves and forest types of Kerala.
11. **Brief study of the following:** Cybernetics, ecological foot print, sustainable development, deep ecology, Gaia hypothesis, conservation ethics, peoples' movements for biodiversity conservation, role of NGOs and educational institutions in biodiversity conservation, trade related IPR, ecotourism.
12. **Climate change and its impacts-** brief study.
13. **Disaster management-** basic aspects.

References

1. Champion H.G. and Seth S.K. A Revised Classification of Forest Types of India. Govt. of India, New Delhi.
2. Gadgil Madhav. Ecological Journeys. Permanent Black, Delhi.
3. Jaiswal P.C. Soil Plant and Water Analysis. Kalyani Publishers, Ludhiana.
4. Krishnamurthy K.V. An Advanced Text Book on Biodiversity Principles and Practice. Oxford IBH.

5. Misra R. Ecology Workbook. Oxford IBH.
6. Odum E.P. and Barrett G.W. Fundamentals of Ecology. Thomson Books, Bangalore.
7. Palmer J.A. Fifty Thinkers on the Environment. Routledge, London.
8. Puri G.S. Indian Forest Ecology. Oxford IBH.
9. Pushpangadan P. and Nair K.S.S. Biodiversity and Tropical Forests- The Kerala Scenario. STEC, Thiruvananthapuram.
10. Sarngdharacharyar. (Translated by Vishnu B.). *Vruksha ayurvedam* Janapriya Pusthakasala, Kottayam.
11. Sivadasan M. and Mohanan K.V. Biodiversity and Ecology: Concepts and Facts. Department of Botany, University of Calicut, Kerala.
12. Speth Gustave James and Haas M. Peter. Global Environmental Governance. Pearson Longman, New Delhi.

BOT4E02 – GENETIC ENGINEERING

Contact Hours per Week: 6

Course Outline

1. Structure of genes in prokaryotes and eukaryotes. Genetic code and codons. Gene expression.
2. Recombinant DNA technology: Tools of rDNA technology, methods of creating rDNA molecules, restriction mapping, isolation and separation of genetic material, southern, northern, western, southwestern and northwestern blotting techniques.
3. Gene transfer techniques in plants- Agrobacterium mediated transfer, gene gun method, electroporation, microinjection, chemical methods.
4. Molecular markers- RAMPO, SSCP, RFLP, RAPD, AFLP, EST markers, Repetitive DNA, Microsatellite and Minisatellite.
5. DNA sequencing- chemical and enzymatic methods. Importance of DNA sequencing.
6. Gel electrophoresis- techniques for visualization and reading sequences.
7. Polymerase Chain Reaction- history, methodology of PCR. Variations from Basic PCR- reverse transcriptase PCR, nested PCR, inverse PCR- applications of PCR.
8. DNA profiling- history, methodology of genetic fingerprinting- applications.
9. Genetic engineering for crop improvement – transgenic plants.
10. Cloning of genes and production of vaccines, drugs, growth hormones and chemicals.
11. Gene therapy- types of gene therapy. Getting transgenes in to patients- viral and non-viral approaches. Success of gene therapy.
12. Abatement of pollution through genetically engineered microorganisms- an emerging approach towards environmental clean-up programmes.
13. Nanotechnology and its applications in genetic engineering.

References

1. Hartl D.L. and Jones E.W. Genetics- Analysis of genes and genome. Jones and Bartlett Publishers.
2. Nicholl Desmond S.T. An Introduction to Genetic Engineering. Cambridge Pub.
3. Brown T.A. Gene Cloning and and DNA Analysis. Blackwell Science Pub.
4. Dubey R.C. A Text Book of Biotechnology. Chand Pub.
5. Singh B.D. Biotechnology. Kalyani Publishers.
6. Walker and Rapley. Molecular Biology and Biotechnology. Panima Pub.
7. Chrispeels M.J. and Sadava D.E. Plants, Genes and Agriculture.
8. Lewin B. Genes. Oxford University Press.
9. Mason A.C. Principles of Gene Manipulation and Genomics.
10. Rissler J. and Mellon M. The Ecological Risks of Engineered Crops. MIT Press, Cambridge.
11. Avice, John C. The Hope, Hype and Reality of Genetic Engineering.
12. McYYan R.P. Genetics and Genetic Engineering. Saras Publications.
13. Narayana L.M. Molecular Biology and Genetic Engineering.
14. Khadpekar N.R. The Age of Nanotechnology. ICFAI University Press, Hyderabad.
15. Nalwa H.R. Encyclopedia of Nanoscience and Technology.

BOT4L07 – PRACTICALS OF ENVIRONMENTAL BIOLOGY, BIODIVERSITY CONSERVATION & GENETIC ENGINEERING

Contact Hours per Week: 6

Course Outline

Environmental Biology and Biodiversity

Practicals

1. Studies on the following and submission of reports: Waste water treatment plant, local environmental peculiarities (such as hillocks and forest patches), wet land ecosystem, alien invasive plants, degraded ecosystem, different forest types, effluent treatment system).
2. Physical and chemical analysis of soil and water: Particle size analysis of soil, estimation of particle density using relative density or volumetric flask; Air capacity analysis of soil by field method; Soil pH analysis of soil using pH meter. Water analysis for pH using pH meter, estimation of BOD by Winkler's method (dark and light bottles).
3. Study of community structure: Charting and mapping of vegetation, Raunkiaer's life forms, biological spectrum, profile diagram (soil).
4. Study of ecological succession: Different types of ecological successions.
5. Visit to an ecological sensitive area and submission of a report.

Genetic Engineering

Practicals

1. Working out problems in genetic engineering.
2. Isolation of plant DNA and its quantification by spectrophotometer.
3. Isolation of plasmid DNA from E. coli.
4. Gel electrophoresis- gel preparation, casting, elution and staining.
5. Visualization of DNA by agarose gel electrophoresis and gel reading.
6. Construction of coding sequence of DNA using amino acid sequence.
7. Visit to a genetic engineering lab and submission of a report.

Audit Courses (To be completed within the first three semesters by the students- Evaluation is 100%

Internal based on Examination /Test (40%) + Seminar / Presentation (30%) + Written assignment (30%) and the marklists are to be forwarded to the university by the end of the third semester)

ACIAEC	Ability Enhancement Course: Scientific Documentation and Report writing	100%	0%	4
AC2PCC	Professional Competency Course: Intellectual Property Rights	100%	0%	4

ACIAEC: Ability Enhancement Course: Scientific Documentation and Report writing

Collection of scientific literature from secondary and primary sources. Preparation of literature reviews and review papers- structure and components Preparation of research papers- structure and components

Scientific conduct, ethics, authorship issues, plagiarism, citation and acknowledgement. Importance of language and effective communication.

Presenting a paper in a scientific seminar- oral and poster presentation Preparation of oral presentations

Preparation of scientific posters

AC2PCC: Professional Competency Course: Intellectual Property Rights

1. Introduction to intellectual property right (IPR)- Concept and kinds. Economic importance. IPR in India and world. IPR and WTO (TRIPS, WIPO).
2. Patents- Objectives, Rights, Patent Act 1970 and its amendments. Procedure of obtaining patents- Working of patents. Infringement.
3. Copyrights- Introduction. Works protected under copyright law. Transfer of Copyright. Infringement. Trademarks- Objectives, Types, Rights. Protection of goodwill. Infringement.
4. Geographical Indications- Objectives, Justification, International Position, Multilateral Treaties, National Level, Indian position.
5. Protection of Traditional Knowledge- Objective, Concept, Holders, Issues concerning, Bio-Prospecting and Bio- Piracy, Alternative ways, Protectability, Traditional knowledge on the International Arena, at

WTO, at National level, Traditional Knowledge Digital Library.

6. Protection of Plant Varieties- Plant Varieties Protection-Objectives, Justification, International Position, Plant varieties protection in India. Rights of farmers, Breeders and Researchers. National gene bank, Benefit sharing. Protection of Plant Varieties and Farmers' Rights Act, 2001.
7. Biotechnology and Intellectual Property Rights- Patenting Biotech Inventions: Objective, Applications, Concept of Novelty, Concept of inventive step, Microorganisms, Moral Issues in Patenting Biotechnological inventions.